

Biological functions of thiamine derivatives: focus on non-coenzyme roles

L Bettendorff*, P Wins

Abstract

Introduction

Thiamine (vitamin B1) is mainly known for its diphosphorylated derivatives, an essential coenzyme in energy metabolism. However, non-coenzyme roles have been suggested for this vitamin for many years. Such roles have remained hypothetical, but recent data from various sources have shed a new light on this hypothesis. First, other phosphorylated thiamine derivatives, most prominently thiamine triphosphate and adenosine thiamine triphosphate, can reach significant levels in *Escherichia coli*, respectively, during amino acid starvation and energy stress. Although much less is known about these compounds in animals, mammalian cells contain a highly specific soluble thiamine triphosphatase controlling cytosolic thiamine triphosphate concentrations. Second, there is now growing evidence in favour of the existence of thiamine-binding proteins with specific roles in the nervous system, possibly in the regulation of neurotransmitter release. Thiamine and some of its synthetic precursors with higher bioavailability have beneficial effects in several models of Alzheimer's disease and may be beneficial for patients suffering from Alzheimer's or Parkinson's diseases. These effects might be related to non-coenzyme roles of thiamine, possibly involving thiamine-binding proteins. The aim of this review was to discuss biological

functions of thiamine derivatives, mainly focusing on non-coenzyme roles.

Conclusion

A hundred years ago, the discovery of thiamine opened the way to the vitamin era of biochemistry, leading to the discovery of the importance of pyruvate oxidation in energy metabolism. This vitamin still has not revealed all of its secrets at a time when metabolomics is emerging as a new powerful tool to refine our knowledge of cellular reactions.

Introduction

Like other B vitamins, thiamine (vitamin B1, Figure 1a) is an indispensable molecule for all known organisms. This is mainly because, in mammalian cells, its diphosphorylated form, thiamine diphosphate (ThDP), is the coenzyme for five key metabolic enzymes (Figure 1b)¹, the most important being mitochondrial pyruvate and oxoglutarate dehydrogenase complexes as well as the cytosolic transketolase. Therefore, it is generally believed that thiamine deficiency leads to decreased oxidative metabolism, which eventually causes cell death. In animals, the brain heavily relies on oxidative metabolism for the synthesis of ATP, making this organ particularly sensitive to thiamine deficiency. In humans, nutritional thiamine deficiency leads to beriberi, a polyneuritic condition, rapidly reversed after thiamine administration. In alcoholics, and also in children, thiamine deficiency can lead to typical selective diencephalic brain lesions² generally referred to as Wernicke-Korsakoff syndrome. The reason why some brain regions are more

sensitive to thiamine deficiency remains unknown³, and it was suggested that this selective vulnerability could be due to a coenzyme-independent role of thiamine or one of its derivatives⁴.

Indeed, in addition to ThDP and free thiamine, several other phosphorylated and adenylated derivatives are observed (Figure 2): thiamine monophosphate (ThMP), thiamine triphosphate (ThTP), adenosine thiamine triphosphate (AThTP) and adenosine thiamine diphosphate (AThDP)^{5,6}. The existence of such forms in many living cells would suggest that they also have some biological role(s). It is indeed worth wondering why the diphosphorylated form of thiamine is the coenzyme, when the monophosphorylated form would do just as well, as is the case for pyridoxal phosphate for instance. It is indeed true that the diphosphate contributes to the binding energy of apoenzymes, but the catalytic properties of thiamine solely rely on the thiazolium ring's ability to lose a proton and form a reactive ylide (Figure 1c). Ylide formation is not influenced by the presence of phosphate groups on the hydroxyethyl arm, and there is no obvious advantage to use ThDP (rather than ThMP or ThTP) as coenzyme.

A recent study emphasises beneficial effects of benfotiamine (a thiamine precursor) in a transgenic mouse model of Alzheimer's disease, although only levels of unphosphorylated thiamine were increased in the brain of the animals. Levels of thiamine-phosphorylated derivatives, including ThDP, were unaffected⁷. Moreover, it was recently suggested that the

* Corresponding author
Email: L.Bettendorff@ulg.ac.be

GIGA-Neurosciences, University of Liège, Liège, Belgium

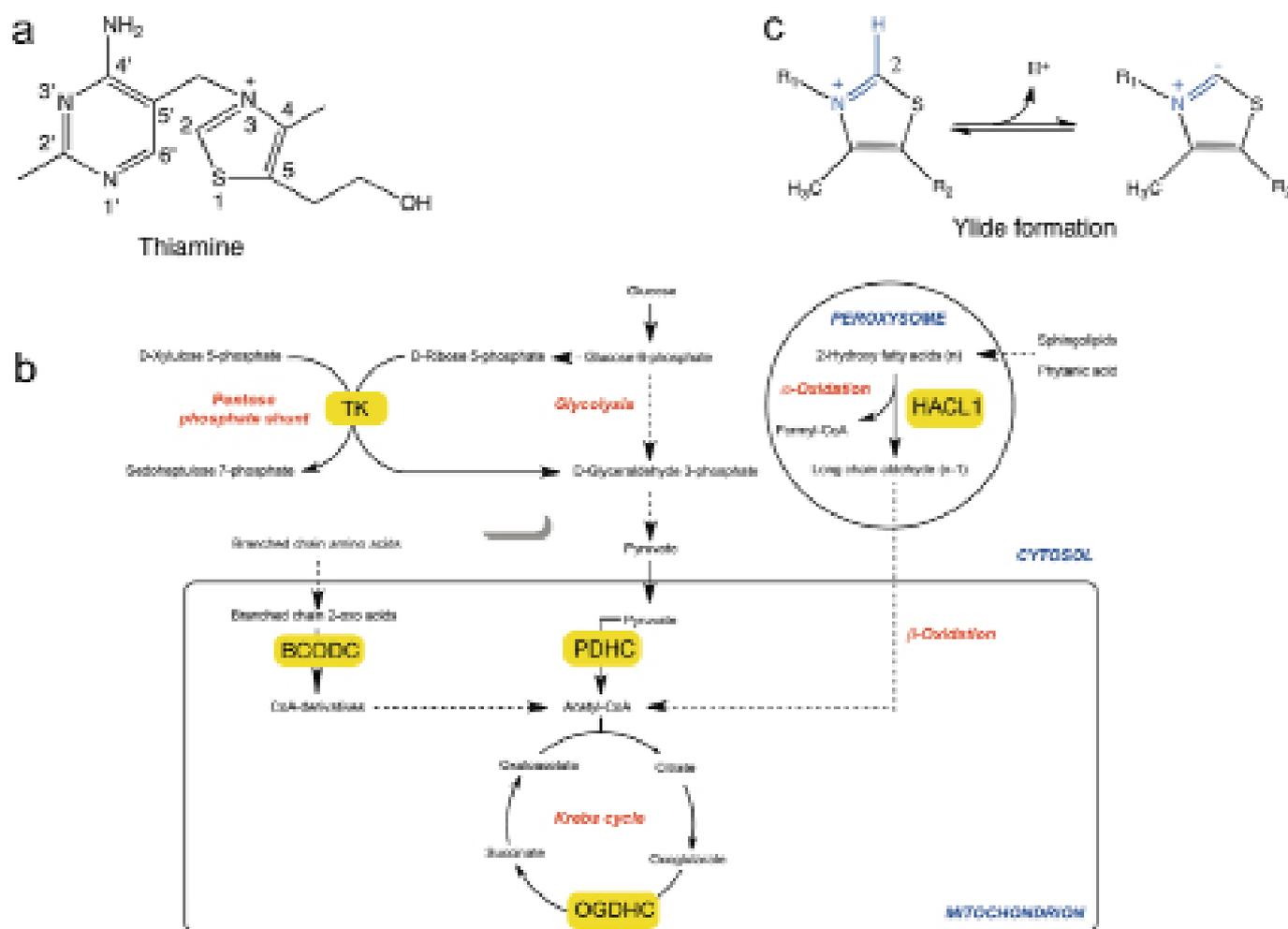


Figure 1: Thiamine diphosphate as a coenzyme. (a) Structural formula of thiamine with both heterocycles numbered according to the usual conventions. (b) Enzyme-catalysed proton loss at the C2 of the thiazolium ring and ylide formation is at the molecular basis of the catalytic properties of thiamine. (c) ThDP-dependent enzymes in a mammalian cell and subcellular localization. TK, transketolase; PDHC, pyruvate dehydrogenase complex; OGDHC, oxoglutarate dehydrogenase complex; BCODC, branched chain 2-oxo acid dehydrogenase complex; HACL1, 2-hydroxyacyl-CoA lyase 1. (Modified from Reference 1).

antinociceptive effects of thiamine in humans and animals could be mediated by the non-phosphorylated form of the vitamin⁸, raising the possibility that free thiamine has pharmacological effects independent of ThDP.

Nearly 20 years ago, we have reviewed data concerning a possible non-coenzyme role of thiamine or its derivatives, particularly in relation to nerve function⁹. Here, we want to critically examine the new data that have been obtained since then.

Discussion

The authors have referenced some of their own studies in this review. These referenced studies have been conducted in accordance with the Declaration of Helsinki (1964), and the protocols of these studies have been approved by the relevant ethics committees related to the institution in which they were performed. All human subjects, in these referenced studies, gave informed consent to participate in these studies. Animal care was also in

accordance with the institution guidelines.

Thiamine derivatives other than ThDP

Thiamine is transported into mammalian cells by specific transporters and immediately phosphorylated to ThDP by cytosolic thiamine pyrophosphokinase (Figure 2). ThDP can then be phosphorylated to ThTP or transformed to adenylated derivatives. However, the most obvious fate for cytosolic-free ThDP is hydrolysis

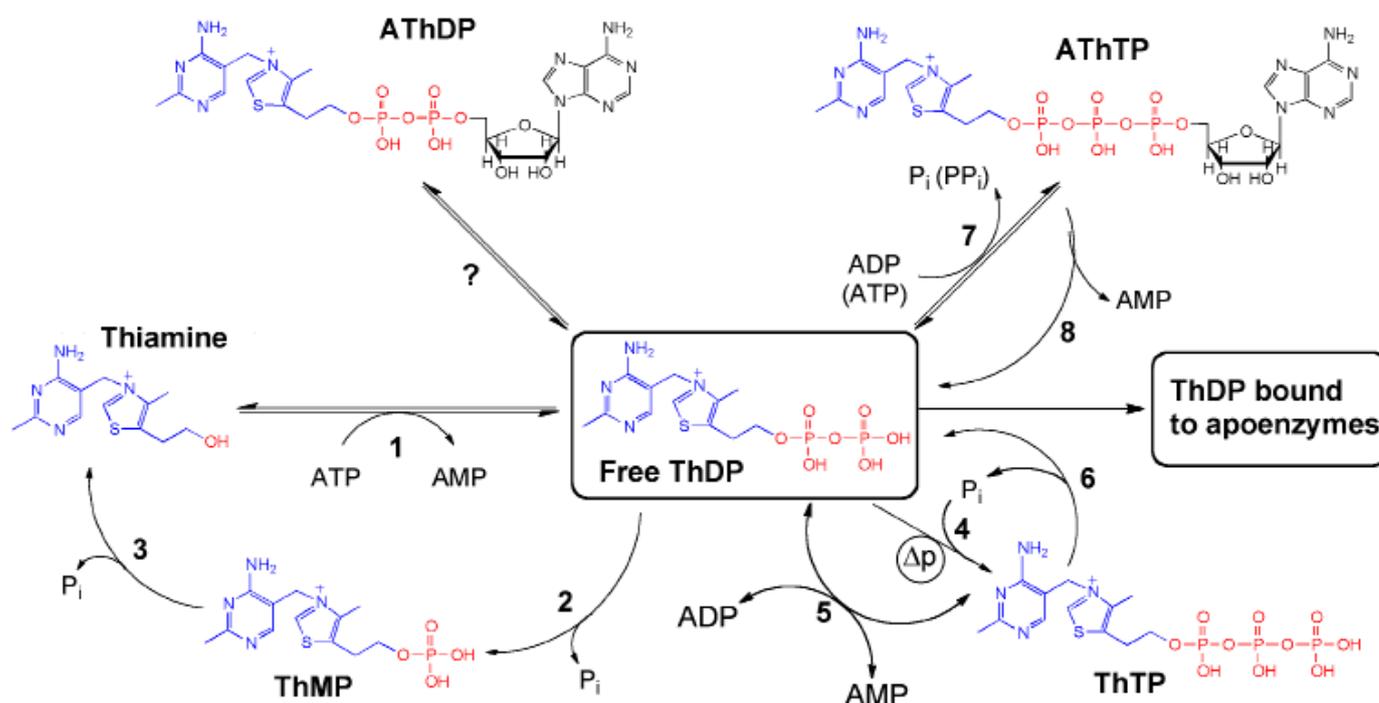


Figure 2: Thiamine derivatives observed in living organisms. (Adapted from References 1 and 63). ThDP is synthesised from thiamine and ATP by thiamine pyrophosphokinase (1). Hydrolysis of ThDP by thiamine pyrophosphatases (2) yields ThMP, which in turn can be hydrolysed to thiamine by thiamine monophosphatases (3). ThDP can be phosphorylated to ThTP by two mechanisms: mitochondrial FoF1-ATP synthase (4) and cytosolic adenylate kinase (5). ThTP can be hydrolysed to ThDP by a very specific cytosolic 25 kDa thiamine triphosphatase (6). ThDP can also be converted to AThTP by a ThDP adenyllyl transferase (7). AThTP can be hydrolysed to ThDP and AMP by a putative AThTP hydrolase (8). AThDP has been shown to exist in prokaryotes and eukaryotes, but its mechanism of synthesis has not yet been demonstrated *in vitro*.

to ThMP, which is recycled to thiamine. No specific enzymes have been identified for the latter reactions, and there is no known role for ThMP. Intracellular ThMP levels are generally much lower than ThDP levels. However, ThMP seems to be excreted, probably by a process involving the reduced folate carrier (RFC1 or SLC19A1)¹⁰, a transporter closely related to thiamine transporters, and it is present in extracellular fluids such as blood plasma, cerebrospinal fluid (CSF) and breast milk.

The case of ThTP

ThTP is a particularly intriguing molecule. It is found in nearly all organisms and is the only known triphosphorylated compound that is not a nucleotide. With two phosphoanhydride bonds, it is an energy-rich compound, and as such it has

been shown to be able to phosphorylate proteins¹¹, although it is not clear whether such phosphorylation is of physiological significance. While ThTP seems to be constitutively synthesised in animal cells, in *Escherichia coli*, it accumulates only in the absence of amino acids and therefore could be a signalling molecule involved in the adaptation to amino acid starvation¹². While it was long thought that ThTP is synthesised by an ATP:ThDP phosphotransferase, the existence of such a mechanism has never been unambiguously demonstrated. It appears now that two ATP-synthesising mechanisms may be diverted towards the synthesis of ThTP: one using adenylate kinase isoform 1 (AK1) (ThDP + ADP \rightleftharpoons ThTP + AMP)¹³ and another via FoF1-ATP synthase by a chemiosmotic mecha-

nism (ThDP + Pi \rightarrow ThTP)^{14–16} in intact *E. coli* cells and isolated brain mitochondria. Interestingly both mechanisms are conserved from bacteria to mammals. However, while the synthesis by adenylate kinase seems to be constitutive and is probably merely a side reaction, the synthesis by FoF1-ATP synthase is strongly dependent on metabolic conditions. While on one hand there is presently no evidence for a specific enzyme involved in ThTP synthesis, on the other hand, mammalian cells contain a highly specific thiamine triphosphatase (ThTPase)^{17–19}. This 25 kDa cytoplasmic protein is a highly efficient ThTPase ubiquitously expressed in adult mammalian tissues. However, it seems to be most abundant in highly differentiated cells while it is hardly detectable in cultured cells, suggesting that

the expression of this enzyme is linked to the degree of cellular differentiation^{20,21}.

It was suggested that ThTPase is a repair enzyme whose role is to remove potentially toxic ThTP produced as a by-product of the above-mentioned reactions²². However, in those organisms where 25 kDa ThTPase is absent (chicken) or catalytically inefficient (fish, pig), cytosolic ThTP indeed accumulates, and in skeletal muscles and electric organs, its levels can even exceed those of ThDP but without apparent toxic effect²¹. It is possible that ThTP has mainly a mitochondrial role, that is, intramitochondrial ThTP synthesised by FoF1-ATP synthase is the physiologically relevant pool, while cytosolic ThTP synthesised by adenylate kinase would be only a by-product of this enzyme activity. In this respect, cytosolic ThTP concentrations might just reflect the abundance of AK1 in the absence of 25 kDa ThTPase.

Adenylated thiamine derivatives

AthTP (or thiaminylated ATP, Figure 2) was first discovered in *E. coli*, where it accumulates in response to carbon starvation or uncoupling^{5,23}. While other B vitamins have long been known to form coenzymes by combination with adenylate (riboflavin in FAD, nicotinic acid in NAD⁺, pantothenic acid in CoA for instance), this was the first time that such a condensation product was demonstrated for thiamine. AthTP exists in small amounts not only in animals and plants (mainly in roots) but also in many cultured mammalian cells²¹. AthTP was shown to be an inhibitor of poly(ADP-ribose) polymerase-1 *in vitro*²⁴. Moreover, small amounts of AthDP were also discovered in various organisms⁶.

Thiamine-binding proteins

We refer here to proteins that specifically bind thiamine or one of its phosphorylated derivatives, but the bound thiamine compound is not supposed

to act as a coenzyme. Likewise, we shall not consider enzymes using thiamine derivatives as substrates (i.e. enzymes involved in the metabolism of phosphoryl derivatives, see Figure 2) nor thiamine transporters.

Several proteins that specifically bind the unphosphorylated form of the vitamin have been described. Some are thiamine-storage proteins, and they were characterised mainly in plant tissues. In mammals, a few thiamine-binding proteins have been described, but their possible roles remain unclear. Such a protein has been purified from rat erythrocytes²⁵. It is a soluble 32 kDa protein binding unphosphorylated thiamine. It is not clear whether it also binds phosphate esters or whether it is specific. The group of Yulia Parkhomenko in Kiev extensively studied thiamine-binding proteins from brain. By affinity chromatography (thiamine covalently bound to a Sepharose 4B matrix), they isolated a thiamine-binding protein from a synaptosomal acetone powder²⁶. This 103–107 kDa protein also binds ThMP and ThTP and to a lesser extent ThDP. The same group later showed that the thiamine-binding activity is mainly associated with synaptic vesicles and synaptosomal membranes²⁶. It was also claimed that this thiamine-binding proteins had ThTPase activity²⁷, but this has not yet been proven using a purified homogenous protein preparation. If this synaptosomal thiamine-binding protein is indeed a membrane-associated membrane protein, it could act as a presynaptic 'thiamine receptor'. There is some evidence that thiamine can act as a neuromodulator at some synapses, regulating neurotransmitter release (see next section). It is also worth pointing out that the antinociceptive effect of thiamine seems to require prostatic acid phosphatase, which could act as or be part of a thiamine receptor⁸.

Synaptosomes prepared from *Torpedo* electric organ are enriched in thiamine and its phosphate esters,

while synaptic vesicles are not, suggesting that they are localised in the axoplasm²⁸. Another study suggested that thiamine is an integral component of synaptosomal membranes²⁹. A role of thiamine in mammalian neuromuscular transmission has also been suggested in other studies³⁰. Taken together, all those data suggest that thiamine may have a specific, coenzyme-independent role in synapses. The existence of ThDP-binding proteins other than apoenzymes using ThDP as coenzyme has long been debated. Cooper and associates claimed that protein-bound ThDP, isolated from a soluble liver fraction, was the substrate for ThTP synthesis³¹, but it was later shown that the only ThDP-binding protein in liver cytosol was transketolase³². In rat brain, Yoshioka et al. described the immunohistochemical localisation of a 68 kDa ThDP-binding protein³³. In this case too, the protein probably corresponds to transketolase as the molecular mass is about the same.

Thiamine in neurotransmitter release

A specific neuroactive role of thiamine in relation to nerve excitability has been postulated as early as the 1940s, and these data have been previously reviewed⁹. While there is presently no convincing evidence that thiamine has physiologically relevant effects on axonal conductance, it has been reported consistently that thiamine (and/or some of its phosphate esters) facilitates neurotransmission in various preparations, probably by potentiation of the release of the neurotransmitters acetylcholine^{28,34,35}, dopamine³⁶ and noradrenaline³⁵. Here, we are exclusively interested in direct (rapid) effects on neurotransmission, as in chronic experiments (for instance, after administration of thiamine for several weeks in animals), it is very difficult to discriminate between putative coenzyme-independent and coenzyme-dependent effects: for instance, increased pyruvate

dehydrogenase activity could lead to increased acetyl-CoA production that in turn could increase acetylcholine synthesis.

In addition to thiamine, several thiamine antimetabolites, the most widely used being pyriothiamine and oxythiamine (Figure 3), have been tested. These structural analogues of thiamine are called antimetabolites, as when administered to animals they produce signs of thiamine deficiency, pyriothiamine acting primarily centrally and oxythiamine acting peripherally as it presumably does not cross the blood-brain barrier. Both compounds competitively inhibit thiamine transport³⁷ and ThDP synthesis by thiamine pyrophosphokinase^{38,39} (although pyriothiamine is more effective).

The fact that they are antimetabolites does not preclude the possibility that they may also act as thiamine agonists when thiamine acts as a non-coenzyme modulator. Indeed, oxythiamine stimulates potassium-evoked acetylcholine release in the presence of Ca^{2+} in isolated brain slices⁴⁰.

These results suggest a coenzyme-independent effect of thiamine on neurotransmitter release, affecting at least three different neurotransmitters (acetylcholine^{28,34,35}, dopamine³⁶ and noradrenaline³⁵) in different preparations ranging from fish electric organ to mammalian brain. This suggests a rather conserved mechanism. Conversely, thiamine deficiency leads to synaptic vesicle dysfunction with decreased release of dopamine⁴¹, glutamate⁴² or acetylcholine⁴³. Moreover, episodes of pyriothiamine-induced thiamine deficiency in the rat lead to a significant reduction in phosphosynapsin I⁴⁴. Although, the animals were treated for 3 weeks with thiamine after appearance of thiamine deficiency symptoms (loss of righting reflex and seizures), the reduction of phosphosynapsin was not reversed. Thus, reduction of phosphosynapsin appears to be an

epigenetic phenomenon that cannot be explained by decrease in ThDP-dependent enzyme activities; indeed, ThDP levels should have been restored by thiamine treatment. It can indeed not be explained by a decrease in ThDP-dependent enzyme activities, as brain thiamine and ThDP levels have presumably been restored. It is thought that phosphorylation of synapsin I leads to a detachment of synapsin from the synaptic vesicles allowing their fusion with the presynaptic membrane and neurotransmitter release. An interesting hypothesis would be that thiamine, directly or indirectly, acts on synapsin I, thereby promoting neurotransmitter release.

This effect could be antagonised by pyriothiamine.

Potential non-coenzyme roles of thiamine and its derivatives are summarised in Figure 4.

Thiamine in stress, diabetes and neurodegenerative diseases

In many instances, beneficial and probiotic effects of thiamine (and/or pharmaceutical preparations of thiamine precursors with higher bioavailability) have been demonstrated. In these cases, we are most likely dealing with pharmacological effects as therapeutic (superphysiological) doses were used. Indeed, under laboratory conditions, either animals or cultured cells are generally in a thiamine-rich

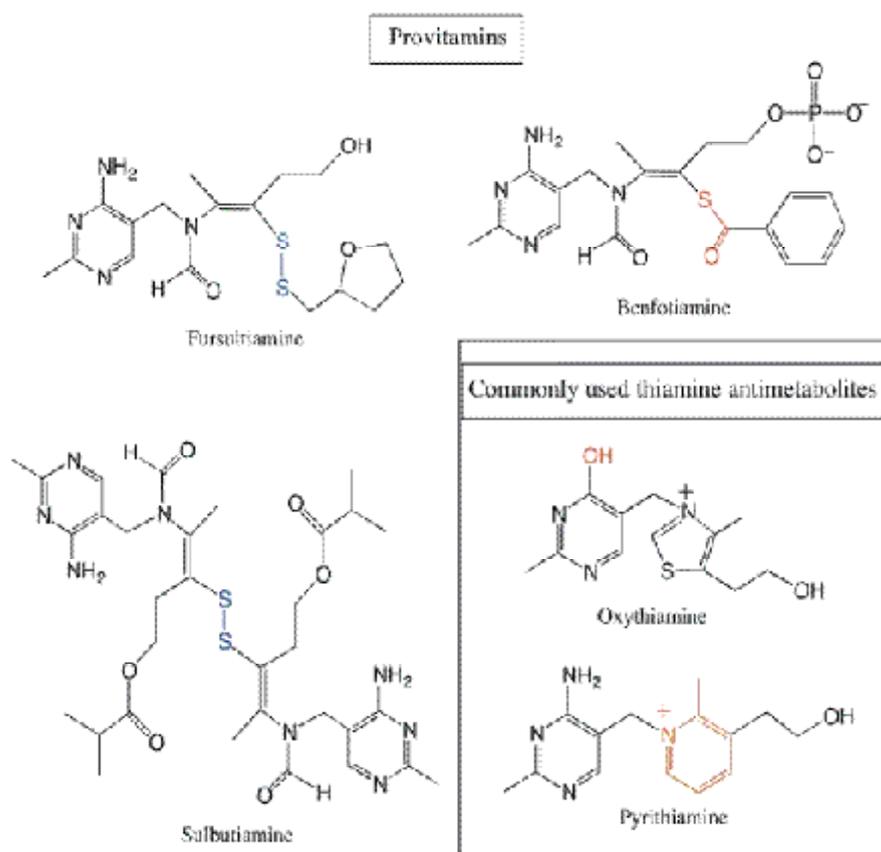


Figure 3: Thiamine provitamins and antimetabolites. Fursultiamine (thiamine tetrahydrofurfuryl disulfide) and sulbutiamine (*O*-isobutyrylthiamine disulfide) are disulfides, while benfotiamine (*S*-benzoylthiamine *O*-monophosphate) is a thioester. The most common thiamine antimetabolites are oxythiamine and pyriothiamine.

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

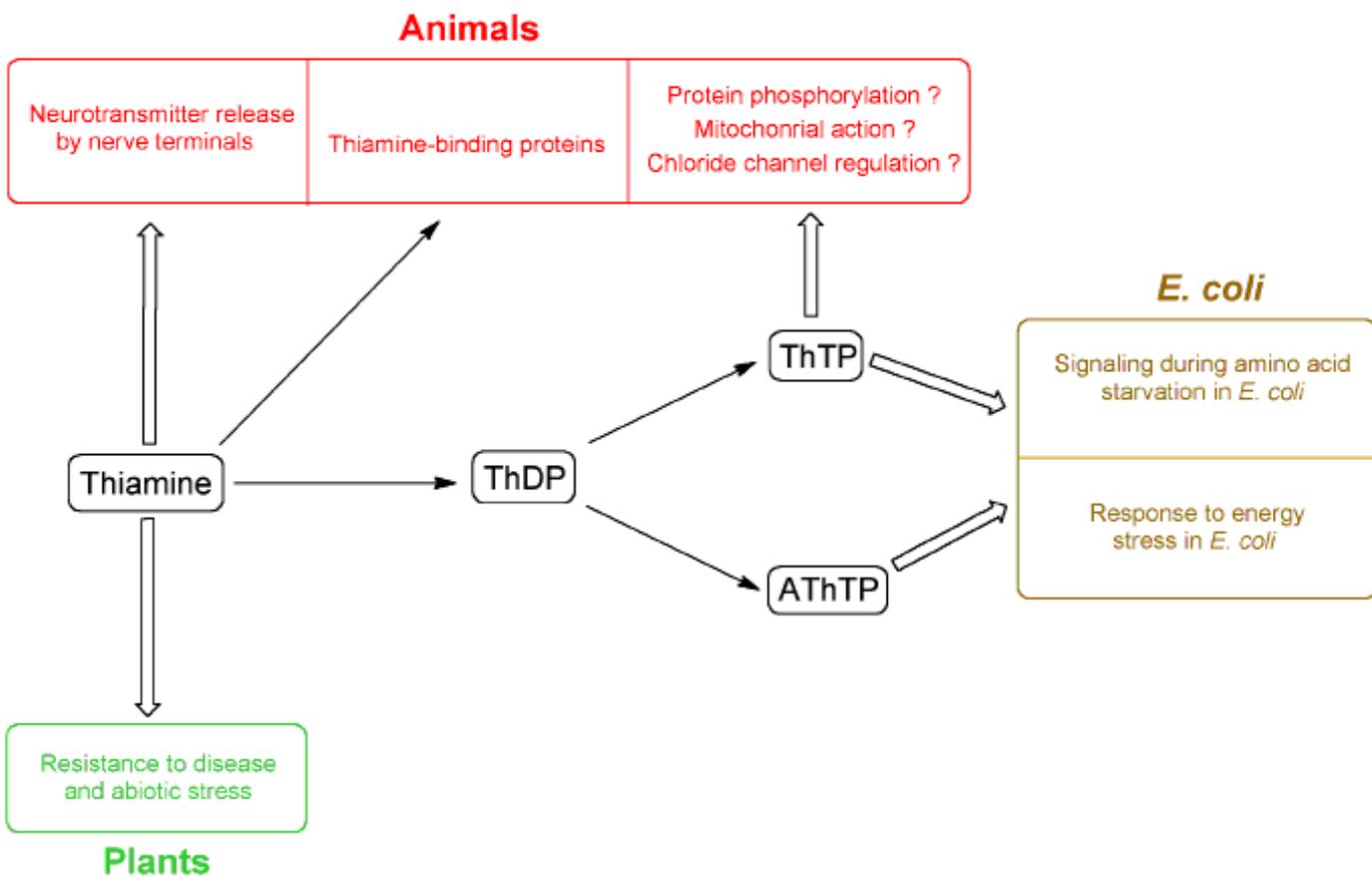


Figure 4: Potential non-coenzyme roles of thiamine and its phosphorylated derivatives. For explanations, see text.

environment: animal chows as well as cell culture media are enriched in vitamins.

According to some reports, thiamine increases disease resistance in plants^{45,46}. Moreover, intracellular thiamine and thiamine phosphate pools are regulated by various stress conditions in *Zea mays* and *Arabidopsis thaliana* seedlings; it was suggested that thiamine is a signalling molecule important for the adaptation to various stress conditions^{47,48}. Interestingly, such a signalling role is assigned to unphosphorylated thiamine in plants, while it should be assigned to ThTP and AThTP in *E. coli*^{5,6,12} (see above). Note that in *Arabidopsis* leaves, ThTP accumulates during withering⁴⁹. Protective effects of thiamine have also been described in mammalian cells: thiamine protects retinal neurons

against glutamate toxicity⁵⁰ and promotes the survival of hippocampal neurons in high cell density culture⁵¹.

Thiamine requires specific transporters to enter cells⁵². As the rate of transport by these transporters is relatively slow, membrane transport is a rate-limiting step in thiamine homeostasis. For that reason, synthetic thiamine precursors were developed. These molecules are either relatively hydrophobic (sulbutiamine, fursultiamine) or are converted to a hydrophobic precursor (benfotiamine) allowing them to cross membranes relatively freely (Figure 3). The general effect of these derivatives is to rapidly increase circulating thiamine to levels higher than those obtained by an equivalent dose of thiamine. It must be emphasised that none of these precursors have ever

been demonstrated to reach the brain parenchyma. They are all converted to thiamine either during the passage from intestine to blood or in the liver. As the blood-brain barrier strongly limits thiamine uptake by the brain (thiamine entry could be limited by a self-exchange), no important increase in brain thiamine levels are observed even with these derivatives^{7,53-55}. It would therefore be interesting to synthesise derivatives that have a half-life sufficiently long to reach significant blood levels to cross the blood-brain barrier.

Nonetheless, thiamine and/or thiamine precursors have been shown to have beneficial effects in diabetes and an animal model of Alzheimer's disease^{7,56,57}. One study has shown improved cognitive functions and a striking decrease in charge of

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

β -amyloid plaques in a mouse model of Alzheimer's disease⁵⁸. This study, however, needs confirmation.

A relationship between thiamine and Parkinson's disease has recently been suggested^{59,60}. It had previously been shown that free thiamine levels are decreased in the CSF of patients with Parkinson's disease compared with control patients⁶¹. Moreover, a very recent preliminary clinical study reported the beneficial effects of thiamine treatment (100–200 mg daily doses of parenteral thiamine) on a limited number of patients⁶². This again needs confirmation.

Conclusion

Thiamine, by the number of its derivatives, is certainly one of the most diverse B vitamins. By virtue of the role of ThDP as coenzyme of several key enzymes, it is involved in nearly all aspects of cell metabolism: energy production, ribose and nucleic acid synthesis, lipid biosynthesis and neurotransmitter synthesis to name only the most important. Therefore, thiamine is particularly important for the nervous system, which is highly sensitive to thiamine deficiency. However, the existence of potential non-coenzyme roles remains a puzzling issue. First, the existence of triphosphorylated derivatives, unable to replace the coenzyme ThDP, is highly suggestive of such roles. ThTP and AThTP may be involved in some signalling processes under specific conditions of cellular stress. Second, thiamine itself, possibly through specific thiamine-binding proteins, may regulate processes such as neurotransmitter release and in plants protect against disease and stress. Although there is still no direct evidence for a physiologically important non-coenzyme role of thiamine, in view of the potential therapeutic interest of thiamine in Alzheimer's and Parkinson's diseases, this may become a key issue in the future.

Acknowledgements

LB is Research Director and PW honorary Research Associate at the 'Fonds de la Recherche Scientifique-FNRS'. This work was supported by grant number 2.4508.10 (LB) from the 'Fonds de la Recherche Fondamentale Collective' (FRFC).

Abbreviations list

AThTP, adenosine thiamine triphosphate; CSF, cerebrospinal fluid; ThDP, thiamine diphosphate; ThMP, thiamine monophosphate; ThTP, thiamine triphosphate; ThTPase, thiamine triphosphatase.

References

- Bettendorff L, Wins P. Biochemistry of thiamine and thiamine phosphate compounds. In: Lane M, DLennarz WJ, editors. Encyclopedia of biological chemistry. Vol. 1. Oxford, UK: Elsevier; 2013. p.202–9.
- Victor M, Adams RD, Collins GH. The Wernicke-Korsakoff syndrome. A clinical and pathological study of 245 patients, 82 with post-mortem examinations. *Contemp Neurol Ser.* 1971;7:1–206.
- Hazell AS, Faim S, Wertheimer G, Silva VR, Marques CS. The impact of oxidative stress in thiamine deficiency: a multifactorial targeting issue. *Neurochem Int.* 2013 Apr;62(5):796–802.
- Cooper JR, Pincus JH. The role of thiamine in nervous tissue. *Neurochem Res.* 1979 Apr;4(2):223–39.
- Bettendorff L, Wirtzfeld B, Makarchikov AF, Mazzucchelli G, Frédéricich M, Gigliobianco T, et al. Discovery of a natural thiamine adenine nucleotide. *Nat Chem Biol.* 2007 Apr;3(4):211–2.
- Frédéricich M, Delvaux D, Gigliobianco T, Gangolf M, Dive G, Mazzucchelli G, et al. Thiaminylated adenine nucleotides. Chemical synthesis, structural characterization and natural occurrence *FEBS J.* 2009 Jun;276(12):3256–68.
- Pan X, Gong N, Zhao J, Yu Z, Gu F, Chen J, et al. Powerful beneficial effects of benfotiamine on cognitive impairment and beta-amyloid deposition in amyloid precursor protein/presenilin-1 transgenic mice. *Brain.* 2010 May;133(Pt 5):1342–51.
- Hurt JK, Coleman JL, Fitzpatrick BJ, Taylor-Blake B, Bridges AS, Vihko P, et al.

Prostatic acid phosphatase is required for the antinociceptive effects of thiamine and benfotiamine. *PLoS One.* 2012;7(10):e48562.

- Bettendorff L. Thiamine in excitable tissues: reflections on a non-cofactor role. *Metab Brain Dis.* 1994 Sep;9(3):183–209.
- Zhao R, Gao F, Goldman ID. Reduced folate carrier transports thiamine monophosphate: an alternative route for thiamine delivery into mammalian cells. *Am J Physiol Cell Physiol.* 2002 Jun;282(6):C1512–7.
- Nghiêm HO, Bettendorff L, Changeux JP. Specific phosphorylation of Torpedo 43K rapsyn by endogenous kinase(s) with thiamine triphosphate as the phosphate donor. *FASEB J.* 2000 Mar;14(3):543–54.
- Lakaye B, Wirtzfeld B, Wins P, Grisar T, Bettendorff L. Thiamine triphosphate, a new signal required for optimal growth of *Escherichia coli* during amino acid starvation. *J Biol Chem.* 2004 Apr 23;279(17):17142–7.
- Shikata H, Koyama S, Egi Y, Yamada K, Kawasaki T. Cytosolic adenylate kinase catalyzes the synthesis of thiamin triphosphate from thiamin diphosphate. *Biochem Int.* 1989 May;18(5):933–41.
- Gigliobianco T, Lakaye B, Makarchikov AF, Wins P, Bettendorff L. Adenylate kinase-independent thiamine triphosphate accumulation under severe energy stress in *Escherichia coli*. *BMC Microbiol.* 2008 Jan;8:16.
- Gangolf M, Wins P, Thiry M, El Moulali B, Bettendorff L. Thiamine triphosphate synthesis in rat brain occurs in mitochondria and is coupled to the respiratory chain. *J Biol Chem.* 2010 Jan;285(1):583–94.
- Gigliobianco T, Gangolf M, Lakaye B, Pirson B, von Ballmoos C, Wins P, et al. An alternative role of FoF1-ATP synthase in *Escherichia coli*: synthesis of thiamine triphosphate. *Sci Rep.* 2013;3:1071.
- Makarchikov AF, Chernikevich IP. Purification and characterization of thiamine triphosphatase from bovine brain. *Biochim Biophys Acta.* 1992 Oct;1117(3):326–32.
- Lakaye B, Makarchikov AF, Antunes AF, Zorzi W, Coumans B, De Pauw E, et al. Molecular characterization of a specific thiamine triphosphatase widely expressed in mammalian tissues. *J Biol Chem.* 2002 Apr;277(16):13771–7.
- Delvaux D, Kerff F, Murty MR, Lakaye B, Czerniecki J, Kohn G, et al. Structural

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

FOR CITATION PURPOSES: Bettendorff L, Wins P. Biological functions of thiamine derivatives: focus on non-coenzyme roles. *OA Biochemistry* 2013 May 01;1(1):10.

- determinants of specificity and catalytic mechanism in mammalian 25-kDa thiamine triphosphatase. *Biochim Biophys Acta*. 2013 Oct;1830(10):4513–23.
20. Czerniecki J, Chanas G, Verlaet M, Bettendorff L, Makarchikov AF, Leprince P, et al. Neuronal localization of the 25-kDa specific thiamine triphosphatase in rodent brain. *Neuroscience*. 2004; 125(4):833–40.
21. Gangolf M, Czerniecki J, Radermecker M, Detry O, Nisolle M, Jouan C, et al. Thiamine status in humans and content of phosphorylated thiamine derivatives in biopsies and cultured cells. *PLoS One*. 2010 Oct;5(10):e13616.
22. Linster CL, Van Schaftingen E, Hanson AD. Metabolite damage and its repair or pre-emption. *Nat Chem Biol*. 2013 Feb;9(2):72–80.
23. Gigliobianco T, Lakaye B, Wins P, El Moulaj B, Zorzi W, Bettendorff L. Adenosine thiamine triphosphate accumulates in *Escherichia coli* cells in response to specific conditions of metabolic stress. *BMC Microbiol*. 2010 May;10:148.
24. Tanaka T, Yamamoto D, Sato T, Tanaka S, Usui K, Manabe M, et al. Adenosine thiamine triphosphate (AThTP) inhibits poly(ADP-ribose) polymerase-1 (PARP-1) activity. *J Nutr Sci Vitaminol (Tokyo)*. 2011;57(2):192–6.
25. Voskoboyev AI, Averin VA. Thiamine-binding protein from rat erythrocytes. *Acta Vitaminol Enzymol*. 1983;5(4):251–4.
26. Parkhomenko YM, Protasova ZS, Yanchiy OR, Khosla K, Donchenko GV. Localization of thiamine-binding protein in synaptosomes from the rat brain. *Neurophysiology*. 2001;33(3):135–9.
27. Parkhomenko IUM, Strokina AA, Pilipchuk Slu, Stepanenko SP, Chekhovskaia LI, Donchenko GV. Existence of two different active sites on thiamine binding protein in plasma membranes of synaptosomes. *Ukr Biokhim Zh*. 2010 Jan-Feb;82(1):34–41.
28. Eder L, Hirt L, Dunant Y. Possible involvement of thiamine in acetylcholine release. *Nature*. 1976;264(5582):186–8.
29. Matsuda T, Cooper JR. Thiamine as an integral component of brain synaptosomal membranes. *Proc Natl Acad Sci U S A*. 1981 Sep;78(9):5886–9.
30. Waldenlind L. Possible role of thiamine in neuromuscular transmission. *Acta Physiol Scand*. 1979 Jan;105(1):1–10.
31. Nishino K, Itokawa Y, Nishino N, Piroso K, Cooper JR. Enzyme system involved in the synthesis of thiamine triphosphate. I. Purification and characterization of protein-bound thiamine diphosphate: ATP phosphoryl transferase. *J Biol Chem*. 1983 Oct;258(19):11871–8.
32. Voskoboev AI, Chernikevich IP. Biosynthesis of thiamine triphosphate and identification of thiamine diphosphate-binding protein of rat liver hyaloplasm. *Biokhimiya*. 1985 Sep;50(9):1421–7.
33. Yoshioka H, Nishino K, Miyake T, Ohshio G, Kimura T, Hamashima Y. Immunohistochemical localization of a new thiamine diphosphate-binding protein in the rat nervous system. *Neurosci Lett*. 1987 Jun;77(1):10–4.
34. Dyatlov VA. Effect of thiamine on the processes responsible for acetylcholine secretion in the frog neuromuscular synapses. *Neurophysiology*. 1994;26:291–8.
35. Romanenko AV, Gnatenko VM, Vladimirova IA. Effect of thiamine on neuromuscular transmission in smooth muscles. *Neurophysiology*. 1994;26(6):370–6.
36. Yamashita H, Zhang YX, Nakamura S. The effects of thiamine and its phosphate esters on dopamine release in the rat striatum. *Neurosci Lett*. 1993 Aug;158(2):229–31.
37. Bettendorff L, Wins P. Mechanism of thiamine transport in neuroblastoma cells. Inhibition of a high affinity carrier by sodium channel activators and dependence of thiamine uptake on membrane potential and intracellular ATP. *J Biol Chem*. 1994 May 20;269(20):14379–85.
38. Liu JY, Timm DE, Hurley TD. Pyriothiamine as a substrate for thiamine pyrophosphokinase. *J Biol Chem*. 2006 Mar; 281(10):6601–7.
39. Voskoboyev AI, Ostrovsky YM. Thiamine pyrophosphokinase: structure, properties, and role in thiamine metabolism. *Ann N Y Acad Sci*. 1982;378:161–76.
40. Hirsch JA, Parrott J. New considerations on the neuromodulatory role of thiamine. *Pharmacology*. 2012;89(1–2):111–6.
41. Mousseau DD, Rao VL, Butterworth RF. Vesicular dysfunction during experimental thiamine deficiency is indicated by alterations in dopamine metabolism. *Eur J Pharmacol*. 1996 Dec;317(2–3):263–7.
42. Lê O, Héroux M, Butterworth RF. Pyriothiamine-induced thiamine deficiency results in decreased Ca²⁺-dependent release of glutamate from rat hippocampal slices. *Metab Brain Dis*. 1991 Sep;6(3):125–32.
43. Jankowska-Kulawy A, Bielarczyk H, Pawelczyk T, Wroblewska M, Szutowicz A. Acetyl-CoA and acetylcholine metabolism in nerve terminal compartment of thiamine deficient rat brain. *J Neurochem*. 2010 Oct;115(2):333–42.
44. Resende LS, Ribeiro AM, Werner D, Hall JM, Savage LM. Thiamine deficiency degrades the link between spatial behavior and hippocampal synapsin I and phosphorylated synapsin I protein levels. *Behav Brain Res*. 2012 Apr;232:421–5.
45. Goyer A. Thiamine in plants: aspects of its metabolism and functions. *Phytochemistry*. 2010 Oct;71(14–15):1615–24.
46. Wang G, Ding X, Yuan M, Qiu D, Li X, Xu C, et al. Dual function of rice OsDR8 gene in disease resistance and thiamine accumulation. *Plant Mol Biol*. 2006 Feb; 60(3):437–49.
47. Rapala-Kozik M, Kowalska E, Ostrowska K. Modulation of thiamine metabolism in *Zea mays* seedlings under conditions of abiotic stress. *J Exp Bot*. 2008;59(15):4133–43.
48. Tunc-Ozdemir M, Miller G, Song L, Kim J, Sodek A, Koussevitzky S, et al. Thiamine confers enhanced tolerance to oxidative stress in Arabidopsis. *Plant Physiol*. 2009 Sep;151(1):421–32.
49. Makarchikov AF, Lakaye B, Gulyai IE, Czerniecki J, Coumans B, Wins P, et al. Thiamine triphosphate and thiamine triphosphatase activities: from bacteria to mammals. *Cell Mol Life Sci*. 2003 Jul;60(7):1477–88.
50. Kaneda K, Kikuchi M, Kashii S, Honda Y, Maeda T, Kaneko S, et al. Effects of B vitamins on glutamate-induced neurotoxicity in retinal cultures. *Eur J Pharmacol*. 1997 Mar;322(2–3):259–64.
51. Geng MY, Saito H, Katsuki H. The effects of thiamine and oxythiamine on the survival of cultured brain neurons. *Jpn J Pharmacol*. 1995 Jul;68(3):349–52.
52. Subramanian VS, Subramanya SB, Said HM. Relative contribution of THTR-1 and THTR-2 in thiamine uptake by pancreatic acinar cells: studies utilizing Slc19a2 and Slc19a3 knockout mouse models. *Am J Physiol Gastrointest Liver Physiol*. 2012 Mar;302(5):G572–8.

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

FOR CITATION PURPOSES: Bettendorff L, Wins P. Biological functions of thiamine derivatives: focus on non-coenzyme roles. *OA Biochemistry* 2013 May 01;1(1):10.

53. Bettendorff L, Weekers L, Wins P, Schoffeniels E. Injection of sulbutiamine induces an increase in thiamine triphosphate in rat tissues. *Biochem Pharmacol.* 1990 Dec;40(11):2557–60.
54. Volvert ML, Seyen S, Piette M, Evrard B, Gangolf M, Plumier JC, et al. Benfotiamine, a synthetic S-acyl thiamine derivative, has different mechanisms of action and a different pharmacological profile than lipid-soluble thiamine disulfide derivatives. *BMC Pharmacol.* 2008 Jun; 8(1):10.
55. Hills JI, Golub MS, Bettendorff L, Keen CL. The effect of thiamin tetrahydrofurfuryl disulfide on behavior of juvenile DBA/2J mice. *Neurotoxicol Teratol.* 2012 Mar;34(2):242–52.
56. Rabbani N, Thornalley PJ. Emerging role of thiamine therapy for prevention and treatment of early-stage diabetic nephropathy. *Diabetes Obes Metab.* 2011 Jul;13(7):577–83.
57. Gibson GE, Hirsch JA, Cirio RT, Jordan BD, Fonzetti P, Elder J. Abnormal thiamine-dependent processes in Alzheimer's Disease. Lessons from diabetes. *Mol Cell Neurosci.* 2013 Jul;55:17–25.
58. Asakura T, Kodera S, Kanda J, Ikeda M. Thiamine-responsive pulmonary hypertension. *BMJ Case Rep.* 2013 Jan.
59. Lu'o'ng KV, Nguyễn LT. Thiamine and Parkinson's disease. *J Neurol Sci.* 2012 Mar;316(1–2):1–8.
60. Costantini A, Pala MI, Compagnoni L, Colangeli M. High-dose thiamine as initial treatment for Parkinson's disease. *BMJ Case Rep.* 2013 Aug;2013.
61. Jiménez-Jimenez FJ, Molina JA, Hernanz A, Fernandez-Vivancos E, de Bustos F, Barcenilla B, et al. Cerebrospinal fluid levels of thiamine in patients with Parkinson's disease. *Neurosci Lett.* 1999 Aug;271(1): 33–6.
62. Luong KV, Nguyen LTH. The beneficial role of thiamine in Parkinson's disease: preliminary report. *J Neurol Res.* 2012; 2(5):211–4.
63. Bettendorff L, Wins P. Thiamin diphosphate in biological chemistry: new aspects of thiamin metabolism, especially triphosphate derivatives acting other than as cofactors. *FEBS J.* 2009 Jun;276(11): 2917–25.