



The analgesic effect of several edible mushrooms

H Wang[†], Y Liu[†], C Han^{*}

Abstract

Introduction

Edible mushrooms, a valuable source of bioactive compounds and nutrients, have been consumed as part of the diet in some countries for thousands of years. They are quite high in protein, carbohydrate and fibre and low in fat content with low trans-isomers of unsaturated fatty acids. In addition, they also have many components such as triterpenes, phenolic compounds, chitosan, eritadenine, sterols (such as ergosterol), triterpenes, etc., which are considered momentous agents for some hitherto unknown healthy properties. Recently, edible mushrooms have become increasingly attractive as functional foods and medicines to treat diseases including cancer, diabetes, inflammation and ache due to the presence of these active components. Pain is an unpleasant sensation, which is a typical response to an untoward event associated with tissue damage, such as injury and inflammation. The aims of this review are to report the positive analgesic effect of several edible mushrooms on pain and its relevant active constituents.

Conclusion

In our review, the edible mushrooms including *Pleurotus pulmonarius*, *Agaricus brasiliensis*, *Agaricus bisporus var. hortensis*, *Agaricus macrosporus*, *Coriolus versicolor* and *Cordyceps sinensis* have been investigated that possess antinociceptive and anti-inflammatory effects owing to their bioactive components such as

β -glucan, agaricoglyceride A, polysaccharopeptide and cordymin as well as other active components. What is more, there are barely any side effects caused by the toxicity of edible mushrooms in vitro and in vivo. However, further research is required with clinical trials and applications.

Introduction

Pain is a physiologically relevant sensation necessary to detect and/or prevent injury; it is sometimes useful to us^{1,2}. Typically, it is a direct response to an untoward event associated with tissue damage, such as injury and inflammation, but severe pain can arise independently of any obvious predisposing cause, or precipitate healing after injury for a relatively long time. It can also occur as a consequence of brain or nerve injury. Pain signalling to the central nervous system is initiated when harmful excitement and primary afferent nociceptive C and A fibres are frequently caused by activation of several types of ionotropic channels and metabotropic receptors^{3,4}. In fact, transient receptor potential and acid-sensing ion channels participate in generating nociceptive signals in response to various specific noxious stimuli⁴⁻⁶. Activity of some of these channels and other proteins implicated in nociceptive signalling pathways can be upregulated by protein kinase C⁷⁻⁹. Thus, pain is generated.

Edible mushrooms are the fleshy and edible fruiting bodies of several species of fungi, typically produced above ground on soil or on its food source¹⁰. They have been used as delicious foods and as healthy nutritional supplements for several centuries. For the Chinese, some mushrooms are especially regarded as medical

substances that boost health and increase longevity, which is attributed to their far-ranging functions, for example antinociceptive, anti-inflammatory, immunity, anti-tumour, ascorbic and so on. Mushrooms are quite high in protein (19–35%) and low in fat. Miles et al. concluded that mushrooms also contain relatively large amounts of carbohydrate and fibre, ranging from 51% to 88% and from 4% to 20%, respectively (dry weight). In addition, mushrooms contain significant amounts of vitamins, namely thiamin, ascorbic acid, riboflavin and vitamin D₂, as well as minerals^{11,12}. In addition to their nutritional value, some mushrooms may also have a medicinal value: anti-tumour, antiviral and hypolipidemic effects have been reported¹¹⁻¹³. They form a huge, but largely untapped powerful source of new pharmaceutical products^{14,15}. They are low-calorie foods with very little fat and are highly suitable for obese persons¹⁶. Their consumption is widespread in China, Japan, Korea, Taiwan, Italy and Spain, among other countries¹⁷⁻¹⁹.

In this review, we intend to discuss the result of research on the antinociception effect of edible mushrooms over the past two decades, emphasising animal studies as well as supporting mechanistic studies. We selected several typical edible fungi which result in pain relief. Our goals are to evaluate the analgesic effect of edible fungus, identify some putative bioactive compounds involved in the effect and stimulate further work in the field.

Discussion

The authors have referenced some of their own studies in this review. The protocols of these studies have been

* Corresponding author

Email: shandongtcmh@163.com

[†] These authors contributed equally to this study.

School of Pharmacy, Shandong University of Traditional Chinese Medicine, Jinan 250355, P. R. China

approved by the relevant ethics committees related to the institution in which they were performed. Animal care was in accordance with the institution guidelines.

Analgesic effect of several edible mushrooms

Oyster mushrooms

Pleurotus is a genus of gilled mushrooms, one of the most widely eaten mushrooms. Species of *Pleurotus* are commonly known as oyster mushrooms and are some of the most commonly cultivated edible mushrooms in the world. The genus *Pleurotus* include edible and medicinal species, most of them being currently commercialised in China²⁰. The fungi are rich in proteins, vitamins, carbohydrates, minerals and dietary fibres. Basidiomycetes have been widely studied over the past 30 years in the light of their polysaccharide composition and therapeutic application^{21,22}.

Pleurotus pulmonarius

Pleurotus pulmonarius, also known as oyster mushroom, is a common edible mushroom consumed worldwide due to its several polysaccharides, besides high amount of proteins, essential amino acids and vitamins^{23,24}. A variety of biological effects have been ascribed to β -glucans, such as anti-inflammatory, antioxidant, anti-tumoural, immunomodulatory and antinociceptive properties²⁵⁻²⁸. There have been some literature research showing that *P. pulmonarius* have analgesic effects²⁹⁻³². Smiderle et al.²⁹ isolated β -glucan (GL) with hot water from the basidiomycete *P. pulmonarius* and found the glucan had potent anti-inflammatory and antinociceptive activities in mice. Animals treated with β -glucan showed a reduction of $85 \pm 5\%$ of writhes induced by acetic acid and the significant inhibition of both the early (neurogenic pain) and the late (inflammatory pain) phases of formalin-induced licking in $43 \pm 5\%$ and $96 \pm 4\%$, respectively.

Then, the results showed that *P. pulmonarius* had notable analgesic and anti-inflammatory effects due to the inhibition of pro-inflammatory cytokines²⁹. The structure of β -D-glucan was characterised using mono- and two-dimensional NMR spectroscopy, methylation analysis and a controlled Smith degradation^{20,29}. And the dates showed that it had a main chain of (1 \rightarrow 6)-linked α -D-galactopyranosyl and 3-O-methyl- α -D-galactopyranosyl units, both of which are partially substituted at O-2 by β -D-mannopyranosyl non-reducing ends (Figure 1).

In order to evaluate the involvement of transient receptor potential (TRP) channels and protein kinase C (PKC) on antinociceptive effect of a (1 \rightarrow 3), (1 \rightarrow 6)-linked β -D-glucan (GL), Baggio et al.³⁰ isolated GL from *P. pulmonarius* to treat it with intraperitoneal administration in mice. In this study, nociceptive responses, induced by intraplantar injections of capsaicin, cinnamaldehyde, menthol, acidified saline and phorbol myristate acetate (PMA), were significantly inhibited by GL. The results demonstrated that GL displayed pronounced systemic antinociceptive properties in chemical models of nociception in mice as a result of the inhibition of PKC ϵ ³⁰. In addition, GL isolated from *P. pulmonarius* could dramatically inhibit acute and neuropathic pain in mice through mechanisms that involve the inhibition of ionotropic glutamate receptors and the interleukin-1 β pathway³¹.

Pleurotus florida

Pleurotus florida, an American oyster mushroom, has been reported to possess antioxidant, immunostimulator, anti-tumour and anti-inflammatory activities^{32,33}. The analgesic and anti-inflammatory activity of *P. florida* was evaluated using a hot plate method, tail flick method, acetic acid-induced writhing, formalin-induced pain and carrageenan-induced inflammation in rats³⁴. Animals treated orally with hydroethanolic extract (HEE) of *P. florida* in a dose-dependent manner were tested for nociceptive response with these methods. Then, results demonstrated *P. florida* exerted excellent analgesic and anti-inflammatory activity in rats on account of myoconstituents like flavonoids, phenolics, polysaccharides and polysaccharopeptides³⁴. Simultaneously, the antinociceptive activity of HEE of *P. florida* is related to the activation of the opioid system.

Pleurotus eous and Pleurotus ostreatus

Pleurotus mushrooms are the second most important mushrooms in terms of production in the world. Furthermore, this species has been of interest to researchers because its phytochemical constituents are similar to those of *P. pulmonarius*, *P. florida*, *Pleurotus eous* and *Pleurotus ostreatus*, which are popularly used in medicines. The ethyl acetate, methanol and aqueous extracts of *P. eous* mushroom were investigated to evaluate

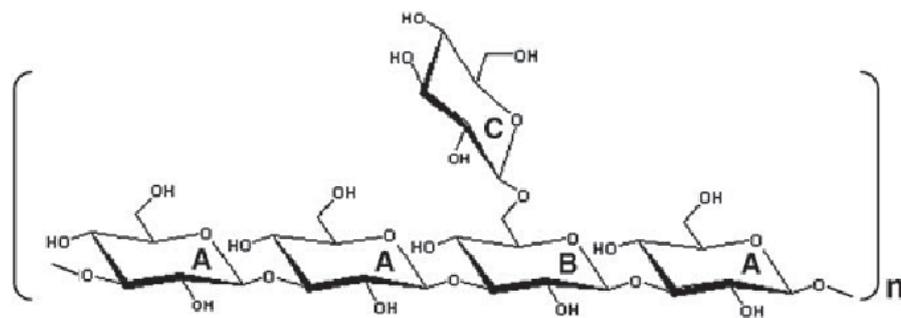


Figure 1: Chemical structure of β -glucan isolated from *Pleurotus pulmonarius*.

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

the analgesic activity using acetic acid-induced writhing, hot-plate, tail immersion and tail-clip tests³⁵. The dates showed that these extracts of *P. eous* produced significant reduction in the number of writhes and markedly raised the pain threshold at different times of observation in comparison with the control ($p < 0.05$). The extracts also caused a notable inhibition of pain in the tail-clip test. Thus, the results of this study revealed that extracts of *P. eous* possessed potent analgesic property and could serve as a base for future drugs³⁵.

Similarly, the antinociceptive potential of *P. ostreatus* was also investigated in rats through the hot-plate, tail-flick and formalin tests³⁶. The reaction times on hot-plate and tail-flick tests were significantly prolonged and pain was significantly suppressed in both phases in the formalin test. The research results showed that *P. ostreatus* had antinociception against neurogenic and continuous inflammatory pain possibly by opioid mechanisms³⁶.

In summary, oyster mushrooms have shown potent antinociceptive and anti-inflammatory properties in several animal model studies and no side effects. However, further studies are needed to investigate the antinociceptive mechanisms in vivo, and human intervention studies of oyster mushrooms alone or in combination with conventional chemotherapy are also demanded to establish efficacy in humans.

Agaricus

Agaricus is the most common eubacterium among the whole macrofungi. The species number admitted by taxonomists is more than 200. It is a large family, which is named *Psalliota Kummer* in the early days, including *Agaricus bisporus*, *Agaricus bitorquis*, *Agaricus blazei*, *Agaricus silvaticus*, *Agaricus macrosporus* and so on. We select three typical species (*A. brasiliensis*, *A. bisporus var. hortensis*

and *A. macrosporus*) in order to elaborate the antinociceptive properties of *Agaricus*.

Agaricus rasilensis and Agaricus bisporis var. hortensis

Fucogalactans from *A. brasiliensis* (EPF-Ab) and *A. bisporus var. hortensis* (EPF-Ah) have antinociceptive action, which is related to their structures. Fucogalactans play a positive role in antinociceptive, anti-inflammatory and anti-sepsis. Besides, they possess activities even after extraction³⁷. The active ingredients are attained by their aqueous extraction and a series of purification. According to methylation analysis³⁸ and GC-MS, Komuraa et al.³⁷ concluded that EPF-Ab (Mw = 19.4×10^3 g/mol) had a (1 → 6)-linked α -D-Galp main-chain partially substituted in O-2 by non-reducing end-units of α -L-Fucp. EPF-Ah (Mw = 31.1×10^3 g/mol) had a similar main-chain with O-2 substitution, but was partially methylated at HO-3, as well as having 2.5% non-reducing end-units of β -D-Gal substitution (Figure 2). Analgesic activity was determined using the hot-plate method, acetic acid-induced writhing, formalin-induced pain in rats and many other classic methods^{37,39}. There are different modes of action among different experiments. Above all, EPF-Ab and EPF-Ah prefer to cure inflammatory nociception and act at

a central and peripheric level. These results showed that the structure determines the function; it is the (1 → 6)-linked α -D-galactopyranosyl main-chain that determines the analgesic property of *A. brasiliensis* and *A. bisporus var. hortensis*. Many articles have reported that a lot of other basidiomycetes' fruiting bodies or cultivated mycelium such as *P. pulmonarius*, *Lentinus edodes*, *Coprinus comatus* and *Hericiium erinaceus*, which have the main-chain, also can inhibit nociception^{20,40-42}.

Agaricus macrosporus

Agaricus macrosporus is another species which has obvious analgesic effect by inhibiting neurolysin. The active ingredient of *A. macrosporus* is agaricoglycerides, which is a new class of fungal secondary metabolites that constitute esters of chlorinated 4-hydroxy benzoic acid and glycerol⁴³. They are produced in cultures of the edible mushroom, which is different from the two species described above. There are seven structures of agaricoglycerides in cultures according to reports, and agaricoglyceride A is the main active principle of the crude extract of *A. macrosporus* (Figure 3). Neurolysin inhibitors are likely to enhance the analgesic properties of neurotensin and/or dynorphin A by inhibiting cleavage and inactivation of these peptides^{44,45}.

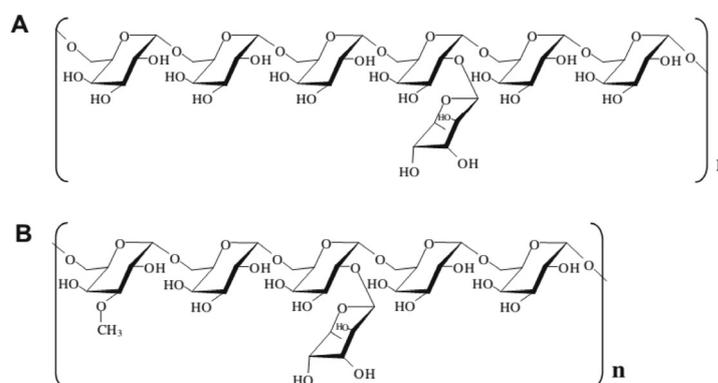


Figure 2: Structure of the fucogalactans EPF-Ab and EPF-Ah, obtained respectively from *A. brasiliensis* (A) and *A. bisporus var. hortensis* (B).

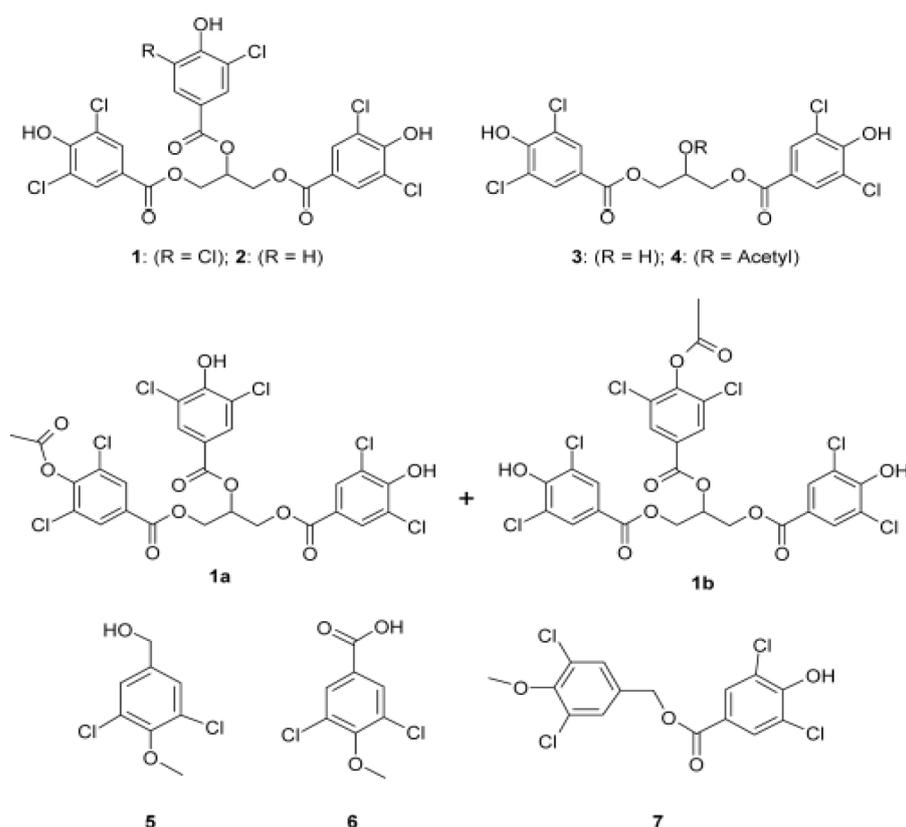


Figure 3: Chemical structures of metabolites isolated from *Agaricus macrosporus*. 1: Agaricoglyceride A; 2: Agaricoglyceride B; 1a/b: Monoacetyl-agaricoglycerides A (isolated as inseparable mixture); 3: Agaricoglyceride C; 4: Agaricoglyceride D; 5: DCMB; 6: 3,5-Dichloro-4-anisic acid; 7: Agaric ester.

The agaricoglyceride A just shows strong activities against neurolysin. Many studies suggested that the production of agaricoglycerides and related metabolites in culture is widespread in the genus *Agaricus*^{46–48} and other genera and families, such as *Triticum*, *Psathyrella*, *Hypholoma*^{49,50} and *Stropharia*, are able to synthesise these aromatic triglycerides.

The genus *Agaricus* generally has an analgesic effect in spite of the kinds of effective ingredients and the different modes of action.

Coriolus versicolor

Coriolus versicolor, also referred to as the turkey-tail mushroom, contains large quantities of β -glucans that act to stimulate the immune system. *Coriolus* can dramatically regenerate and rejuvenate the body. Its most

active medicinal components are biological response modifiers called protein-bound polysaccharides. Both extracellular and intracellular polysaccharopeptides of *C. versicolor* are physiologically active as biological response modifiers. The best known commercial polysaccharopeptide preparations of *C. versicolor* are polysaccharopeptide Krestin (PSK) and polysaccharopeptide (PSP). What is more, PSP and PSK have been investigated for possessing anticancer, anti-inflammatory and antinociceptive activities and immunopotentialiation^{51–55}.

The PSP from the mushroom *C. versicolor* has immunomodulatory and anti-tumour activities, which has been used as a drug for cancer patients^{56,57}. Ng et al.⁵⁴ studied the analgesic activity of PSP in the

hot-plate test upon intraperitoneal administration to ICR mice and found that its analgesic activity would add to its attribute as an immunomodulatory and anti-tumour drug. Pain response in mice was significantly suppressed after receiving an intraperitoneal injection of *C. versicolor* PSP. Then, it was demonstrated that *C. versicolor* PSP possesses analgesic activity, which is beneficial for cancer patients as an immunomodulatory and anti-tumour drug⁵⁴. In addition to this, one study shows that the analgesia produced by PSP is mediated by IL-2, which is activated by PSP and interacts with IL-2 receptors in the mediobasal hypothalamus (MBH)⁵⁵. Rats were divided at random into different groups to test and the degree of analgesic effect of PSP was evaluated by the pain threshold (mA) or by the percentage increase from baseline pain threshold. The experiment results demonstrated that the analgesia appeared only 1 h after PSP administration and began to decrease after another hour. The phenomenon suggested that PSP-produced analgesia might be mediated by some intermediary substances activated by PSP⁵⁵.

On the whole, *C. versicolor* as a medicinal mushroom is widely used to treat cancer and enhance the immune system. Its PSK and PSP are the main bioactive components possessing anti-tumour, antimicrobial, hepato-protective and analgesic effects. Even so, the clinical applications need further study.

Cordyceps

Among the whole macrofungi family, *Cordyceps* is a special genus of entomogenous fungi that forms a fruiting body mainly on pupae or larvae⁵⁸. The most typical species is *Cordyceps sinensis* (CS, caterpillar fungus). It is named Dong-Chong-Xia-Cao in Chinese, which translates as winter worm and summer grass. CS has many pharmacological actions such as modulation of immune response,

inhibition of tumour growth, induction of cell apoptosis, antinociception and so on^{59–62}. The effective constituent of antinociception in CS is cordymin, a peptide purified from its culture and fruiting bodies. Some studies have shown that cordymin inhibits the acetic acid-induced abdominal constriction in mice in a dose-dependent manner, which shows that cordymin had a peripheral antinociceptive effect. In addition to the results of the hot-plate test which is used for the assessment of the central antinociceptive effect, cordymin significantly inhibited the reaction time to thermal stimuli^{63,64}. In brief, cordymin has antinociceptive effect in both peripheral and central aspects. Cordymin-1, cordymin-2 and cordymin-4 inhibited neurolysin in a dose-dependent manner and neurolysin has been reported to have analgesic properties in animal models⁶⁵. As a result, cordymin is a potent anti-inflammatory and analgesic medicine and CS is an effective analgesic.

Others

Not only the four genera described above but also many others have the effect of analgesia. Lu et al.⁶⁶ concluded that the dry matter of culture broth (DMCB) of *Termitomyces albuminosus* in submerged culture, its crude saponin extract (CSE) and crude polysaccharide extract (CPE) inhibited the mouse ear swelling by 61.8%, 79.0% and 81.6%, respectively. Then the data illustrated that *T. albuminosus* possessed the analgesic activity owing to saponins and polysaccharides, which are its major active constituents⁶⁶. One study, designed by Park et al., demonstrated that the methanol extract from *Inonotus obliquus* had analgesic activity due to the inhibition of iNOS and COX-2 expression via the down-regulation of NF- κ B binding activity⁶⁷. In addition, Kim et al.⁶⁸ found that the EtOH extract of *Phellinus linteus* (PLE) could significantly reduce the numbers of writhing induced by acetic

Edible mushrooms	Bioactive components/extracts of analgesic effect	References
<i>Pleurotus pulmonarius</i>	β -Glucans	[25–32]
<i>Pleurotus florida</i>	Hydroethanolic extract	[34]
<i>Pleurotus eous</i>	Methanol and aqueous extracts	[35]
<i>Agaricus brasiliensis</i>	Fucogalactan (EPF-Ab)	[37]
<i>Agaricus bisporus var. hortensis</i>	Fucogalactan (EPF-Ah)	[37]
<i>Agaricus macrospores</i>	Agaricoglycerides	[43–48]
<i>Coriolus versicolor</i>	Polysaccharopeptides	[51–57]
<i>Cordyceps</i>	Cordymin	[63–65]
<i>Termitomyces albuminosus</i>	Crude saponin extract Crude polysaccharide extract	[66]
<i>Inonotus obliquus</i>	Methanol extract	[67]
<i>Phellinus linteus</i>	EtOH extract	[68]
<i>Lactarius rufus</i>	Soluble β -D-glucan	[69]
<i>Grifola frondosa</i>	Agaricoglycerides	[70]

acid in mice. The results indicated that PLE had potent antinociceptive effect, which might be mediated by its anti-inflammatory action⁶⁸. Moreover, Ruthes et al.⁶⁹ studied and found that *Lactarius rufus* had the anti-inflammatory and antinociceptive potential of their polysaccharides evaluated using the formalin model. Soluble β -D-glucan isolated from fruiting bodies of *L. rufus* produced potent inhibition of inflammatory pain caused by formalin when compared with the insoluble one⁶⁹. Furthermore, a recent study stated that *Grifola frondosa* has important and antinociceptive effects in acetic acid-induced pain and formalin-induced inflammatory pain at the dose level of 500 mg/kg in mice⁷⁰. Therefore, *G. frondosa* may be used as an alternative medicine for inflammatory pain.

Conclusion

Edible mushrooms have been widely used in some cultures as traditional

medicines to treat diseases including diabetes and cancer, and to stimulate the immune system. Pain is intuitive for feelings of these diseases such as cancer, inflammation and injuries. As a result, the analgesic effects of edible fungi have a wide range of applications. The active components in many mushrooms with analgesic effects are very clear. In our review, edible mushrooms including *P. pulmonarius*, *A. brasiliensis*, *A. bisporus var. hortensis*, *A. macrosporus*, *C. versicolor* and CS have been investigated that possessed antinociceptive and anti-inflammatory effects owing to their bioactive components such as β -glucan, agaricoglyceride A, polysaccharopeptide and cordymin as well as other active components (Table 1). What is more, there are barely any side effects caused by toxicity of edible mushrooms in vitro and in vivo. However, further research is required with clinical trials and applications.

Competing interests: none declared. Conflict of interests: none declared. All authors contributed to the conception, design, and preparation of the manuscript, as well as read and approved the final manuscript. All authors abide by the Association for Medical Ethics (AME) ethical rules of disclosure.



Abbreviations list

CPE, crude polysaccharide extract; CS, *Cordyceps sinensis*; CSE, crude saponin extract; DMCB, dry matter of culture broth; HEE, hydro-ethanolic extract; MBH, mediobasal hypothalamus; PKC, protein kinase C; PMA, phorbol myristate acetate; PSK, polysaccharopeptide Krestin; PSP, polysaccharopeptide; TRP, transient receptor potential.

Acknowledgement

This work was supported by the Foundation of Ji'nan Science and Technology Development Program (201302055).

References

- Julius D, Basbaum AI. Molecular mechanisms of nociception. *Nature*. 2001 Sep;413(6852):203–10.
- Velazquez KT, Mohammad H, Sweitzer SM. Protein kinase C in pain: involvement of multiple isoforms. *Pharmacol Res*. 2007 Jun;55(6):578–89.
- Scholz J, Woolf CJ. Can we conquer pain? *Nat Neurosci*. 2002 Nov;5:1062–7.
- Tominaga M. Nociception and TRP channels. *Handb Exp Pharmacol*. 2007; 179:489–505.
- Caterina MJ, Julius D. Sense and specificity: a molecular identity for nociceptors. *Curr Opin Neurobiol*. 1999 Oct;9(5):525–30.
- McCleskey EW, Gold MS. Ion channels of nociception. *Annu Rev Physiol*. 1999; 61:835–56.
- Baron A, Deval E, Salinas M, Lingueglia E, Voilley N, Lazdunski M. Protein kinase C stimulates the acid-sensing ion channel ASIC2a via the PDZ domain-containing protein PICK1. *J Biol Chem*. 2002 Dec; 277(52):50463–8.
- Ferreira J, Triches KM, Medeiros R, Calixto JB. Mechanisms involved in the nociception produced by peripheral protein kinase c activation in mice. *Pain*. 2005 Sep;117(1–2):171–81.
- Premkumar LS, Raisinghani M, Pingle SC, Long C, Pimentel F. Down-regulation of transient receptor potential melastatin 8 by protein kinase C-mediated dephosphorylation. *J Neurosci*. 2005 Dec;25(49): 11322–9.
- Chang ST, Miles PG. Mushrooms: cultivation, nutritional value, medicinal effect, and environmental impact. Boca Raton, Florida: CRC Press; 1989.pp.4–6.
- Miles P, Chang S-T. Mushroom biology. Concise basics and current developments. Singapore, New Jersey, London, Hong Kong: World Scientific; 1997.
- Breene WM. Nutritional and medicinal value of specialty mushrooms. *J Food Protect*. 1990;53:883.
- Johl PP, Sodhi HS, Dhanda S, Kapoor S. Mushrooms as medicine—a review. *J Plant Sci Res*. 1995–6;11–12:73.
- Obodai M, Vowotor KA. Performance of different strains of *Pleurotus* species under Ghanaian conditions. *J Food Technol Afr*. 2002;1:98–100.
- Tong HB, Xia FG, Feng K. Structural characterization and in vitro antitumor activity of a novel polysaccharide isolated from the fruiting bodies of *Pleurotus ostreatus*. *Bioresour Technol*. 2009 Feb; 100(4):1682–6.
- Garcha HS, Khann PK, Soni GL. Nutritional importance of mushroom. In: Chang ST, Buswell JA, Chin S, editors. Mushroom biology and mushroom products. Hong Kong: The Chinese University Press; 1993.pp.227–35.
- Chang ST. The development of the mushroom industry in China, with a note on possibilities for Africa. *Acta Ed Fung*. 2005;12:3–19.
- Moreno-Rojas R, Díaz-Valverde MA, Arroyo BM, González TJ, Capote CJB. Mineral content of gurumelo (*Amanita ponderosa*). *Food Chem*. 2004;85:325–30.
- Manzi P, Gambelli L, Marconi S, Vivanti V, Pizzoferrato L. Nutrients in edible mushrooms: an inter-species comparative study. *Food Chem*. 1999;65:477–82.
- Smiderle FR, Olsen LM, Carbonero ER, Marcon R, Baggio CH, Freitas CS, et al. A 3-O-methylated mannogalactan from *Pleurotus pulmonarius*: structure and antinociceptive effect. *Phytochemistry*. 2008 Nov;69(15):2731–6.
- Smith JE, Sullivan R, Rowan N. The role of polysaccharides derived from medicinal mushrooms in cancer treatment programs: current perspectives (review). *Int J Med Mushrooms*. 2003;5:217–23.
- Zhang M, Cui SW, Cheung PCK, Wang Q. Antitumor polysaccharides from mushrooms: a review on their isolation process, structural characteristics and antitumor activity. *Trends Food Sci Technol*. 2007; 18:4–19.
- Wasonga CG, Okoth SA, Mukuria JC, Omwandho CO. Mushroom polysaccharide extracts delay progression of carcinogenesis in mice. *J Exp Ther Oncol*. 2008;7(2):147–52.
- Yatsuzuka R, Nakano Y, Jiang S, Ueda Y, Kishi Y, Suzuki Y, et al. Effect of Ushiratake (*Pleurotus pulmonarius*) on sneezing and nasal rubbing in BALB/c mice. *Biol Pharm Bull*. 2007 Aug;30(8):1557–60.
- Mizuno T, Hagiwara T, Nakamura T, Ito H, Shimura K, Sumiya T, et al. Antitumor activity and some properties of water-soluble polysaccharides from “Himematsutake,” from the fruiting body of *Agaricus blazei* Murill. *Agric Biol Chem*. 1990;54:2889–96.
- Toklu HZ, Sener G, Jahovic N, Uslu B, Arbak S, Yegen BÇ. β -Glucan protects against burn-induced oxidative organ damage in rats. *Int Immunopharmacol*. 2006 Feb;6(2):156–69.
- Dore CMPG, Azevedo TCG, Souza MCR, Rego LA, Dantas JCM, Silva FRF, et al. Anti-inflammatory, antioxidant and cytotoxic actions of β -glucan-rich extract from *Geastrum saccatum* mushroom. *Int Immunopharmacol*. 2007 Sep;7(9):1160–9.
- Zhang M, Cui SW, Cheung PCK, Wang Q. Antitumor polysaccharides from mushrooms: a review on their isolation process, structural characteristics and antitumor activity. *Trends Food Sci Technol*. 2007;18:4–19.
- Smiderle FR, Olsen LM, Carbonero ER, Baggio CH, Freitas CS, Marcon R, et al. Anti-inflammatory and analgesic properties in a rodent model of a (1 \rightarrow 3),(1 \rightarrow 6)-linked β -glucan isolated from *Pleurotus pulmonarius*. *Eur J Pharmacol*. 2008 Nov;597(1–3):86–91.
- Baggio CH, Freitas CS, Marcon R, de Paula Wernera MF, Rae GA, Smiderle FR, et al. Antinociception of β -d-glucan from *Pleurotus pulmonarius* is possibly related to protein kinase C inhibition. *Int J Biol Macromol*. 2012 Apr;50(3):872–7.
- Baggio CH, Freitas CS, Martins DF, Mazzardo L, Smiderle FR, Sasaki GL, et al. Antinociceptive effects of (1/3), (1/6)-linked β -glucan isolated from *Pleurotus pulmonarius* in models of acute and neuropathic pain in mice: evidence for a role for glutamatergic receptors

Competing interests: none declared. Conflict of interests: none declared. All authors contributed to the conception, design, and preparation of the manuscript, as well as read and approved the final manuscript. All authors abide by the Association for Medical Ethics (AME) ethical rules of disclosure.

- and cytokine pathways. *J Pain*. 2010 Oct; 11(10):965–71.
32. Roy SK, Das D, Mondal S, Maiti D, Bhunia B, Maiti TK, et al. Structural studies of an immunoenhancing water-soluble glucan isolated from hot water extract of an edible mushroom, *Pleurotus florida*, cultivar Assam Florida. *Carbohydr Res*. 2009 Dec;344(18):2596–601.
33. Jose N, Ajith TA, Janardhanan KK. Methanol extract of the oyster mushroom, *Pleurotus florida*, inhibits inflammation and platelet aggregation. *Phytother Res*. 2004 Jan;18(1):43–6.
34. Ganeshpurkar A, Rai G. Experimental evaluation of analgesic and anti-inflammatory potential of Oyster mushroom *Pleurotus florida*. *Indian J Pharmacol*. 2013 Jan–Feb;45(1):66–70.
35. Suseem SR, Saral MA, Reddy NP, Gregory M. Evaluation of the analgesic activity of ethyl acetate, methanol and aqueous extracts of *Pleurotus eous* mushroom. *Asian Pac J Trop Med*. 2011 Feb;4(2):117–20.
36. Vasudewa NS, Abeytunga DT, Ratna-sooriya WD. Antinociceptive activity of *Pleurotus ostreatus*, an edible mushroom in rats. *Pharm Biol*. 2007;45:533–40.
37. Komuraa DL, Carbonerob ER, Grachera AHP, Baggio CH, Freitas CS, Marcon R, et al. Structure of *Agaricus spp.* fucogalactans and their anti-inflammatory and antinociceptive properties. *Bioresour Technol*. 2010 Aug;101(15):6192–9.
38. Ciucanu I, Kerek F. A simple and rapid method for the permethylation of carbohydrates. *Carbohydr Res*. 1984;131:209–17.
39. Ruthes AC, Rattmann YD, Malquevicz-Paiva SM, Carbonero ER, Córdova MM, Baggio CH, et al. *Agaricus bisporus* fucogalactan: structural characterization and pharmacological approaches. *Carbohydr Polym*. 2013 Jan;92(1):184–91.
40. Carbonero ER, Gracher AHP, Komura, et al. *Lentinus edodes* heterogalactan: antinociceptive and anti-inflammatory effects. *Food Chem*. 2008 Dec;111(3):531–7.
41. Fan J, Zhang J, Tang Q, Liu Y, Zhang A, Pan Y. Structural elucidation of a neutral fucogalactan from the mycelium of *Coprinus comatus*. *Carbohydr Res*. 2006 Jul;341(9):1130–4.
42. Zhang A, Zhang J, Tang Q, Jia W, Yang Y, Liu Y, et al. Structural elucidation of a novel fucogalactan that contains 3-O-methyl rhamnose isolated from the fruiting bodies of the fungus, *Hericium erinaceus*. *Carbohydr Res*. 2006 Apr; 341(5):645–9.
43. Stadler M, Hellwig V, Mayer-Bartschmid A, Denzer D, Wiese B, Burkhardt N. Novel analgesic triglycerides from cultures of *Agaricus macrospores* and other basidiomycetes as selective inhibitors of neurolysin. *J Antibiot*. 2005 Dec;58(12):775–86.
44. Vincent B, Dive V, Yiotakis A, Smadja C, Maldonado R, Vincent JP, et al. Phosphorus-containing peptides as mixed inhibitors of endopeptidase 3. 4. 24. 15 and 3. 4. 24.16: effect on neurotensin degradation *in vitro* and *in vivo*. *Brit J Pharmacol*. 1995 Jul;115(6):1053–63.
45. Jiráček J, Yiotakis A, Vincent B, Checler F, Dive V. Development of the first potent and selective inhibitor of the zinc endopeptidase neurolysin using a schematic approach based on combinatorial chemistry of phosphinic peptides. *J Biol Chem*. 1996 Aug;271(32):19608–11.
46. Hirotani M, Sai K, Nagai R, Hirotani S, Takayanagi H, Yoshikawa T. Blazeispirane and protoblazeispirane derivatives from the cultured mycelia of the fungus *Agaricus blazei*. *Phytochemistry*. 2002 Nov;61(5): 589–95.
47. Hirotani M, Sai K, Hirotani S, Yoshikawa T. Blazeispirols B, C, E and F, des-A-ergostane-type compounds from the cultured mycelia of the fungus *Agaricus blazei*. *Phytochemistry*. 2002 Mar;59(5):571–7.
48. Zapf S, Anke T, Dasenbrock H, Steglich W. Antifungal metabolites from *Agaricus sp.* 89139. *Bioengineering*. 1:92.
49. Hautzel R, Anke H. Screening of ascomycetes and basidiomycetes for plant growth regulating substances: introduction of the gibberellic acid induced de novo synthesis of hydrolytic enzymes in embryoless seeds of *Triticum aestivum* as test system. *Z Naturforsch*. 1990;45C:1093–8.
50. Swarts HJ, Verhagen FJM, Field JA, Wijnberg JBPA. Trichlorinated phenols from *Hypholoma elongatum*. *Phytochemistry*. 1998;49:203–6.
51. Cho HJ, Shim MJ, Choi EC, Kim BK. Studies of constituents of higher fungi of Korea LVII. Comparison of various anti-tumor constituents of *Coriolus versicolor*. *Kor J Mycol*. 1988;16:162–74.
52. Sakagami H, Aoki T, Simpson A, Tanuma SI. Induction of immunopotential activity by a protein-bound polysaccharide, PSK (review). *Anticancer Res*. 1991 Mar–Apr;11(2):993–1000.
53. Li XY, Wang JF, Zhu PP, Liu L, Ge JB, Yang SX. Immune enhancement of a polysaccharides peptides isolated from *Coriolus versicolor*. *Acta Pharmacol Sinica*. 1990 Nov;11(6):542–5.
54. Ngand TB, Chan WY. Polysaccharopeptide from the mushroom *Coriolus versicolor* possesses analgesic activity but does not produce adverse effects on female reproductive or embryonic development in mice gen. *Pharmacology*. 1997; 29(2):269–73.
55. Shan G, Hui-Qin Z, Wei-Ping Y, Qi-Zhang Y, Yi Z, Zhen-Lun G, et al. Involvement of interleukin-2 in analgesia produced by *Codolus versicolor* polysaccharide peptides. *Acta Pharmacologica Sinica*. 1998 Jan;19(1):67–70.
56. Liao ML, Zhao JM. Stage 11 clinical tests of PSP in the treatment of lung cancer. In: Yang QY, Kwok CY, editors. *Proceedings of international symposium on PSP*. Shanghai, China: Fudan University Press; 1993. pp.243–56.
57. Shi JH, Chen T, Lian ZR. Clinical research of the effect of PSP on the immunological function of stomach cancer patients during operation and chemotherapy. In: Change ST, editor. *Proceedings of international symposium on PSP*. Shanghai, China: Fudan University Press; 1993. pp.232–40.
58. So-Young W, Eun-Hee P. Anti-inflammatory and related pharmacological activities of cultured mycelia and fruiting bodies of *Cordyceps militaris*. Seoul, South Korea: College of Pharmacy, Sookmyung Women's University.
59. Kuo YC, Tsai WJ, Wang JY, Chang SC, Lim CY, Shiao MS. Regulation of bronchoalveolar lavage fluids cell function by the immunomodulatory agents from *Cordyceps sinensis*. *Life Sci*. 2001 Jan;68(9):1067–82.
60. Kuo YC, Lin CY, Tsai WJ, Wu CL, Chen CF, Shiao MS. Growth inhibitors against tumor cells in *Cordyceps sinensis* other than cordycepin and polysaccharides. *Cancer Invest*. 1994;12(6):611–5.
61. Bok JW, Lerner L, Chilton J, Klingeman HG, Towers GH. Antitumor sterols from the mycelia of *Cordyceps sinensis*. *Phytochemistry*. 1999 Aug;51(7):891–8.
62. Yang LY, Huang WJ, Hsieh HG, Lin CY. H1-A extracted from *Cordyceps sinensis*



suppresses the proliferation of human mesangial cells and promotes apoptosis, probably by inhibiting the tyrosine phosphorylation of Bcl-2 and Bcl-XL. *J Lab Clin Med.* 2003 Jan;141(1):74–83.

63. Collier HO, Dinneen JC, Johnson CA, Schneider C. The abdominal constriction response and its suppression by analgesic drugs in the mouse. *Br J Pharmacol Chemother.* 1968;32:295–310.

64. Qian GM, Pan GF, Guo JY. Anti-inflammatory and antinociceptive effects of cordymin, a peptide purified from the medicinal mushroom *Cordyceps sinensis*. *Nat Prod Res.* 2012;26(24):2358–62.

65. Tyler BM, Cusack B, Douglas CL, Souder T, Richelson E. Evidence for

additional neurotensin receptor subtypes: neurotensin analogues that distinguish between neurotensin-mediated hypothermia and antinociception. *Brain Res.* 1998 May;792(2):246–52.

66. Lu YY, Ao ZH, Lu ZM, Xu HY, Zhang XM, Dou WF, et al. Analgesic and anti-inflammatory effects of the dry matter of culture broth of *Termitomyces albuminosus* and its extracts. *J Ethnopharmacol.* 2008 Dec;120(3):432–6.

67. Park YM, Won JH, Kim YH, Choi JW, Park HJ, Lee KT. In vivo and in vitro anti-inflammatory and anti-nociceptive effects of the methanol extract of *Inonotus obliquus*. *J Ethnopharmacol.* 2005 Oct;101(1–3):120–8.

68. Kim SH, Song YS, Kim SK, Kim BC, Lim CJ, Park EH. Anti-inflammatory and related pharmacological activities of the n-BuOH subfraction of mushroom *Phellinus linteus*. *J Ethnopharmacol.* 2004 Jul;93(1):141–6.

69. Ruthes AC, Carbonero ER, Córdova MM, Baggio CH, Santos ARS, Sasaki GL, et al. *Lactarius rufus* (1→3),(1→6)-β-d-glucans: structure, antinociceptive and anti-inflammatory effects. *Carbohydr Polym.* 2013 Apr;94(1):129–36.

70. Han C, Cui B. Pharmacological and pharmacokinetic studies with agaricoglycerides, extracted from *Grifola frondosa*, in animal models of pain and inflammation. *Inflammation.* 2012 Aug;35(4):1269–75.

Competing interests: none declared. Conflict of interests: none declared. All authors contributed to the conception, design, and preparation of the manuscript, as well as read and approved the final manuscript. All authors abide by the Association for Medical Ethics (AME) ethical rules of disclosure.

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)